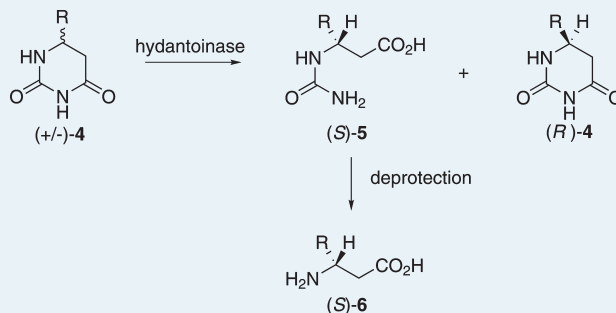


Enzyme-Catalyzed Enantioselective Hydrolysis of Dihydrouracils as a Route to Enantiomerically Pure β -Amino AcidsMaeve O'Neill,[†] Bernhard Hauer,[‡] Nina Schneider,[‡] and Nicholas J. Turner^{*,†}[†]School of Chemistry, University of Manchester, Manchester Interdisciplinary Biocentre, Manchester, M1 7DN, U.K.[‡]BASF AG, Ludwigshafen, Germany

ABSTRACT: The hydantoinase from *Vigna angularis* has been shown to catalyze the hydrolysis of a range of racemic 6-substituted dihydrouracils to yield the corresponding *N*-carbamoyl-(*S*)- β -amino acids and unreacted (*R*)-dihydrouracils. High enantioselectivity ($E > 100$) was achieved in cases that the C-6 substituent was an aryl group. Subsequent treatment of the *N*-carbamoyl derivatives with nitrous acid yielded the free β -amino acid.

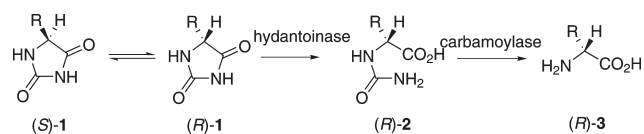
KEYWORDS: hydantoinase, β -amino acids, enantioselective, biocatalysis, dihydrouracil, *Vigna angularis*



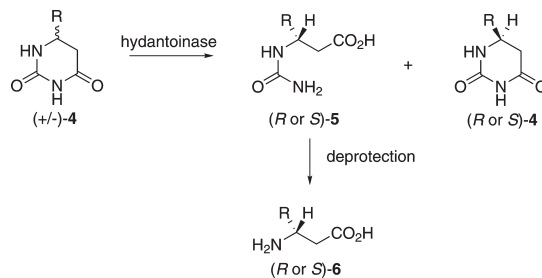
Enantiomerically pure β -amino acids are valuable building blocks for novel therapeutic agents that possess a wide range of biological activity.^{1–3} Although a number of biocatalytic routes have been developed for their preparation, no single method has emerged as being universally applicable.⁴ Recent approaches have been based upon the use of transaminases,⁵ lipases,^{6,7} and aminopeptidases.^{8,9} The isomerization of α - to β -amino acids using aminomutases offers a potentially attractive route, although this approach is currently hampered by the need to separate equilibrium mixtures of α -/ β -amino acids.^{10,11}

The use of hydantoinases for the enantioselective hydrolysis of racemic 5-substituted hydantoins **1** to their corresponding *N*-carbamoyl derivatives **2** is well established (Scheme 1) and has been developed to the stage that commercial processes now operate at scale for the production of specific D-(*R*)-amino acids **3** using this technology.¹² A key aspect of these processes is the in situ racemization of the unreacted enantiomer (*S*)-**1** together with carbamoylase catalyzed hydrolysis of (*R*)-**2**, leading to a dynamic kinetic resolution (DKR) reaction.

In contrast, the possibility of carrying out enantioselective hydrolysis of 6-substituted dihydrouracils **4** to their corresponding *N*-carbamoyl derivatives **5** as a route to β -amino acids **6** has received very little attention (Scheme 2). May et al.¹³ described the use of a hydantoinase from *Arthrobacter aurescens* for the hydrolysis of dihydrouracil (**4**; R=H) and subsequently it was reported¹⁴ that this hydantoinase could be applied to the resolution of 6-phenyldihydrouracil (**4**; R=Ph), although poor enantioselectivity and low reaction rates relative to 5-phenylhydantoin (**1**; R=Ph) were observed. In a separate study, a Japanese group¹⁵ reported e.e.'s of up to 51% for the hydrolysis of 6-phenyldihydrouracil using a *Bacillus* sp.; higher selectivities (up to 93% e.e.) were obtained with substrates containing 6-alkyl, rather than 6-aryl, substituents. To assess the viability of this approach to β -amino acids, we decided to examine the hydrolysis

Scheme 1. Hydantoinase/Carbamoylase Catalyzed DKR of Racemic 5-Substituted Hydantoins **1** To Yield D-(*R*)-Amino Acids (**3**)

Scheme 2. Hydantoinase Catalyzed Hydrolysis of Racemic 6-Substituted Dihydrouracils



of a range of 6-substituted dihydrouracils (\pm)-**4** using the commercially available hydantoinase from *Vigna angularis*.¹⁶

The required 6-substituted dihydrouracil substrates **4a–i** were prepared by one of two alternative routes (Scheme 3). In method A,^{17,18} urea was heated with the appropriate cinnamic acid derivative **7** at 190 °C for 2–4 h, followed by

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Scheme 3. Synthesis of Racemic 6-Substituted Dihydrouracils

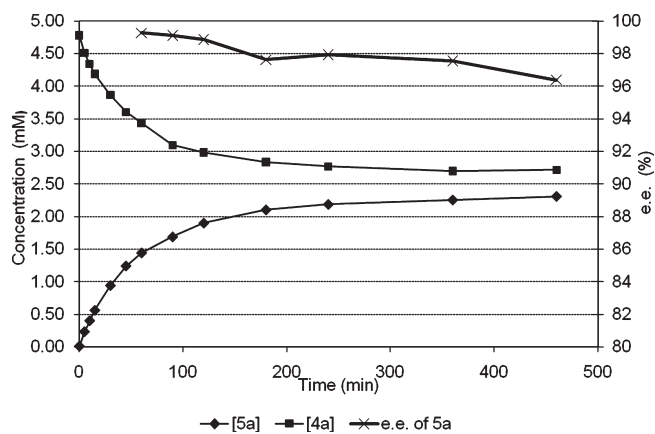
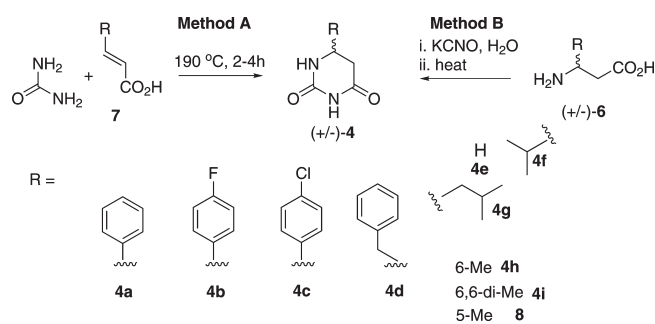


Figure 1. Progress of hydantoinase catalyzed hydrolysis of (\pm)-4a to (S)-5a showing composition of reaction mixture and e.e.

recrystallization of the product, to yield 4a–i in moderate yields of up to 46%. Method B¹⁹ involved treatment of the corresponding racemic β -amino acid 6 with potassium cyanate to generate the *N*-carbamoyl derivative, followed by heating in concentrated HCl to effect cyclization to give 4a–i in good overall yields. Method B, which is known to proceed without racemization, was also used to prepare enantiomerically pure (S)-4a and (S)-5a, by using enantiomerically pure (S)-6a as the starting material.

Initially, the hydantoinase-catalyzed hydrolysis of racemic 6-phenyldihydrouracil (4a: R=Ph) was studied as a model system. Reactions were carried out at a substrate concentration of 5 mM in TRIS buffer and monitored by reversed-phase chiral HPLC, which allowed simultaneous determination of both the extent of conversion in the reaction and also the e.e. of unreacted dihydrouracil 4a and the *N*-carbamoyl derivative 5a. The following observations were noted:

- The *V. angularis* hydantoinase was found to be highly selective (e.e. up to 98%) for the *S*-enantiomer of 4a with an *E* value >100. The absolute configurations of both the product (S)-5a and unreacted substrate (R)-4a were assigned by comparison with authentic samples. (S)-5a is the expected enantiomer based upon the known selectivity of the *V. angularis* hydantoinase for (R)-hydantoins 1 (Scheme 1).
- Despite the high *S* enantioselectivity observed, conversions did not proceed to 50% as expected, even after prolonged reaction times. All reactions gave an equilibrium ratio of 4a:5a = 54:46 after 6 h with an e.e. for 5a of ~97% (Figure 1).

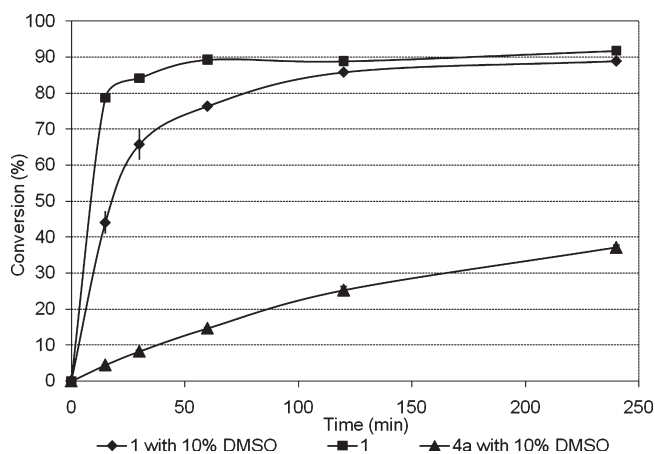
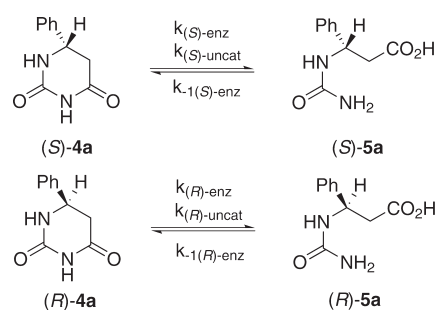
Scheme 4. Kinetic Scheme for Hydrolysis of (\pm)-4a with *V. angularis* Hydantoinase

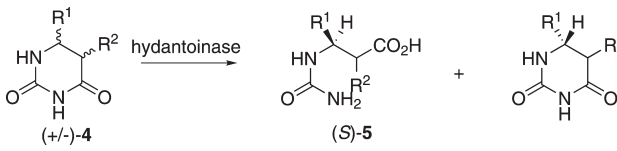
Figure 2. Relative rates of hydrolysis of 1 and 4a using *V. angularis* hydantoinase in the presence and absence of DMSO.

- To further probe the equilibrium issue, the reverse reaction, namely, hydantoinase-catalyzed cyclization of *N*-carbamoyl- β -phenylalanine 5a, was investigated. At pH 7.5 with (\pm)-5a, ~5% conversion to 4a was observed after 6 h, confirming the equilibrium position at this pH. As expected, the cyclization was found to be highly *S*-selective with no appreciable cyclization of the *R* enantiomer ($k_{-1S-enz} > k_{-1R-enz}$). No cyclization occurred in the absence of the hydantoinase.
- In contrast to hydantoins, appreciable background hydrolysis of 4a occurred with a strong pH and buffer dependency. At pH 7 in TRIS buffer, only 0.5% hydrolysis was observed, rising to 20% at pH 9. The hydantoinase was found to exhibit maximum activity at pH = 7.5 with respect to hydrolysis of (\pm)-4a.

The diagram in Scheme 4 summarizes the overall kinetic scheme for the hydrolysis of (\pm)-4a to 5a. In this process $k_{S-enz} \gg k_{R-enz}$ and $k_{-1S-enz} > k_{-1R-enz}$ with $k_{S-uncat} = k_{R-uncat}$. Although reversibility has also been demonstrated for hydantoinase-catalyzed hydrolysis of hydantoins,²⁰ this reaction has an appreciable rate only at low pH, in contrast to cyclization of 5a.

Direct comparison with 5-phenylhydantoin (1: R=Ph) as substrate revealed that 4a was hydrolyzed with ~10% of the rate when the reaction was carried out in DMSO (Figure 2).

The substrate specificity of the hydantoinase was then examined with respect to a range of 6-substituted dihydrouracils 4a–j (Table 1).

Table 1. Hydrolysis of 4i–4j with *V. angularis* Hydantoinase


substrate	R ¹	R ²	rel. rate	E
4a	C ₆ H ₅	H	7	>200
4b	<i>p</i> -F-C ₆ H ₄	H	15	>100
4c	<i>p</i> -Cl-C ₆ H ₄	H	nd	>100
4d	C ₆ H ₅ CH ₂	H	79	0
4e	H	H	100	
4f	<i>i</i> -Pr	H	46	3
4g	<i>i</i> -Bu	H	55	2
4h	Me	H	52	5
4i	(Me) ₂	H	0	
4j	H	Me	23	

The reaction was found to be highly enantioselective ($E > 100$), provided that the substrate contained an aryl group as the C-6 substituent (4a–c). Replacement of phenyl- (4a) with benzyl- (4d) resulted in a high rate of hydrolysis but no apparent selectivity. In cases that the C-6 substituent is an alkyl group (4f–h), the reactions proceed quickly but with low selectivity ($E_R = 2–5$). Relocating the substituent at the C-5 rather than C-6 position (4j) led to a reduced rate of hydrolysis with no apparent selectivity.

Finally, a preparative scale reaction was carried out at 20 mM substrate concentration. After 4 h, the conversion of (\pm)-4a to the product (*S*)-5a reached 46% with 97% e.e. Subsequent extraction of the reaction mixture with 2-butanol gave (*S*)-5a in 42% yield based on (\pm)-4a. Cleavage of the *N*-carbamoyl group with nitrous acid yielded (*S*)- β -phenylalanine 6a in 79% yield. An attempt to remove the *N*-carbamoyl group by treatment of (*S*)-5a with a commercially available carbamoylase²¹ was not successful, in contrast to a previous report using the carbamoylase from *Agrobacterium tumefaciens*.²²

In summary, we have demonstrated that racemic 6-substituted dihydrouacils can be enantioselectively hydrolyzed by the hydantoinase from *V. angularis*. In cases that the substituent is an aryl group, the selectivity is very high ($E > 100$), thereby providing a new and practical method for the preparation of this class of enantiomerically pure β -amino acids.

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REFERENCES

- Weiner, B.; Szymanski, W.; Janssen, D. B.; Minnaard, A. J.; Feringa, B. L. *Chem. Soc. Rev.* **2010**, *39*, 1656–1691.
- Enantioselective synthesis of beta amino acids*, 2nd Ed.; Juaristi, E., Soloshonok, V. A., Eds.; Wiley-VCH Verlag GmbH: Weinheim, 2005.
- Cardillo, G.; Tomasini, C. *Chem. Soc. Rev.* **1996**, *25*, 117–128.
- Liljeblad, A.; Kanerva, L. T. *Tetrahedron* **2006**, *62*, 5831–5854.
- Savile, C. K.; Janey, J. M.; Mundorff, E. C.; Moore, J. C.; Tam, S.; Jarvis, W. R.; Colbeck, J. C.; Krebber, A.; Fleitz, F. J.; Brands, J.; Devine, P. N.; Huisman, G. W.; Hughes, G. J. *Science* **2010**, *239*, 305–309.

- Tasnádi, G.; Forró, E.; Fülöp, F. *Org. Biomol. Chem.* **2010**, *8*, 793–799.
- Shakeri, M.; Engstroem, K.; Sandstroem, A. G.; Baeckvall, J.-E. *ChemCatChem* **2010**, *2*, 534–538.
- Heck, T.; Seebach, D.; Osswald, S.; ter Wiel, M. K. J.; Kohler, H.-P. E.; Geueke, B. *ChemBioChem* **2009**, *10*, 1558–1561.
- Grundmann, P.; Fessner, W.-D. *Adv. Synth. Catal.* **2008**, *350*, 1729–1735.
- Turner, N. J. *Curr. Opin. Chem. Biol.* **2011**, *15*, 234–240.
- Verkuijl, B. J. V.; Szymański, W.; Wu, B.; Minnaard, A. J.; Janssen, D. B.; de Vries, J. G.; Feringa, B. L. *Chem. Commun.* **2010**, *46*, 901–903.
- May, O.; Nguyen, P. T.; Arnold, F. H. *Nat. Biotechnol.* **2000**, *18*, 317–320.
- May, O.; Siemann, M.; Pietzsch, M.; Kiess, M.; Mattes, R.; Sylđatk, C. *J. Biotechnol.* **1998**, *61*, 1–13.
- Servi, S.; Sylđatk, C.; Vielhauer, O.; Tessaro, D. Poster Communication in *Biotrans* (3-8.7.2005); Delft, The Netherlands, 2005.
- Yamada, M.; Kamyama, N.; Yasohara, Y.; Takahashi, S.; Hasegawa, J. *Jpn. Kokai Tokkyo Koho*, Ed. Japan; JP 06261787, 1994.
- Partially purified D-hydantoinase from *V. angularis* (H4028, 468 U_g¹⁻) and the immobilized hydantoinase from *Escherichia coli* (53765, 50 U_g¹⁻) were obtained from Sigma.
- Lee, C. K.; Shim, J. Y. *B. Kor. Chem. Soc.* **1991**, *12*, 343–347.
- Vuano, B. M.; Peroni, O. I.; Cabaleiro, M. C. *J. Chem. Res.* **2000**, *7*, 318–320.
- Posner, T. *Ber. Dtsch. Chem. Ges.* **1905**, *38*, 2316–2325.
- Ohishi, T.; Nanba, H.; Sugawara, M.; Izumida, M.; Honda, T.; Mori, K.; Yanagisawa, S.; Ueda, M.; Nagashima, N.; Inoue, K. *Tetrahedron Lett.* **2007**, *48*, 3437–3440.
- The D-carbamoylase (DCRB-01) was obtained from Codexis.
- Martinez-Gomez, A. I.; Martinez-Rodriguez, S.; Pozo-Dengra, J.; Tessaro, D.; Servi, S.; Clemente-Jimenez, J. M.; Rodriguez-Vico, F.; Las Heras-Vazquez, F. J. *Appl. Environ. Microb.* **2009**, *75*, 514–520.